Sprouts of the broccoli cultivar Everest contained 130-fold more inducer potential (units/g fresh weight) than mature vegetables. The inducer activity in broccoli was significantly higher than in daikon.

Example 5 INDUCER POTENTIAL OF BROCCOLI SPROUT EXTRACTS

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Inducer potential of a series of water extracts of 3-day old broccoli sprouts of the cultivar Saga were Plants were prepared by first surface determined. sterilizing seeds of Brassica oleracea variety italica (broccoli) cultivar Saga by a 1 min treatment in 70% ethanol, followed by 15 min in 1.3% sodium hypochlorite containing approximately 0.001% Alconox detergent. Seeds were grown in sterile plastic containers at a density of approximately 8 seeds/cm2 for 72 hours on a 0.7% agar support that did not contain added nutrients. environment was carefully controlled with broad spectrum fluorescent lighting, humidity and temperature control (16 hours light, 25°C / 8 hours dark, 20°C).

Plants were rapidly and gently collected from the surface of the agar to minimize glucosinolate hydrolysis by endogenous myrosinase released upon plant wounding. Sprouts (approximately 25 mg fresh wt/sprout) were gently harvested and immediately and rapidly plunged into approximately 3 volumes of boiling water in order to inactivate endogenous myrosinase as well as to extract glucosinolates and isothiocyanates from the plant tissue. Water was returned to a boil and maintained at a rolling boil for 3 min. The sprouts were then either strained from the boiled infusion [tea, soup] or homogenized in it, and the residue then removed by filtration or centrifugation.

Data in Table 3 represent both homogenates and infusions. Preparations were stored at -20°C until assayed. Inducer potential of plant extracts, prepared

Exhibit 4-C

as described above, was determined as described in Definitions section above.

TABLE 3 Inducer Potentials of Hot Water Extracts of 3-Day Saga Broccoli Sprouts

	_			
		EXTRACT NO.	units/g fresh weight	
		1	500,000	
103	,00 _	2	370,000	
		3	455,000	
	10	4	333,000	
C		5	435,000	
		6	333,000	
U U		7	625,000	
ā		8	250,000	
2 2	15	9	313,000	
		10	357-, 000	
		11	370,000	
Ō		12	370,000	
		13	217,000	
	20	14	222,000	
		15	1,000,000	
		16	714,000	
		17	435,000	
		18	1,250,000	
	25	19	263,000	
		AVERAGE	464,000 ± 61,600 S.E.M.	
		<u> </u>		

Some variability in the amount of Phase 2 enzyme-inducer potential was detected. High levels of Phase 2 enzyme-inducer potential, however, were consistently observed.

Example 6

HOT WATER BROCCOLI EXTRACTS TREATED WITH DAIKON MYROSINASE

QR activity in a hot water broccoli extract increased in the presence of a vegetable source of myrosinase. An aqueous extraction of 3-day old sprouts of broccoli cultivar Saga grown on water agar, in which myrosinase was inactivated by boiling for 3 min, was divided into 6 different 150 ml aliquots. Nine-day old daikon sprouts, a rich source of the enzyme myrosinase, were added to this cooled infusion in amounts equivalent to 0, 5, 9, 17, 29 and 40% (w/w) of the broccoli. QR activity, as determined in the Definition section, of the control extracts containing 0% daikon was 26,300 units/gram fresh weight while QR activity of the extracts that had received daikon as a source of myrosinase ranged from 500,000 to 833,000 units/gram fresh weight of broccoli. Accordingly, myrosinase present in the daikon sprouts, increased the QR activity in the broccoli extract greater than 19-fold.

Example 7

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GLUCORAPHANIN AND GLUCOERUCIN ARE THE PREDOMINANT GLUCOSINOLATES IN HOT WATER EXTRACTS OF BROCCOLI (CULTIVAR SAGA) SPROUTS

Paired Ion Chromatography (PIC). Centrifuged hot water extracts of 3-day-old broccoli (cultivar Saga) sprouts were subjected to analytical and preparative PIC on a reverse phase C18 Partisil ODS-2 HPLC column in ACN/H₂O (1/1, by vol.) with tetraoctylammonium (TOA) bromide as the counter-ion. Only three well-separated peaks were detected: peak A eluted at 5.5 min, B at 11.5

min, and C at 13 min at a molar ratio [A:B:C] of ca. 2.5 : 1.6 : 1.0 (monitored by UV absorption at 235 nm), and they disappeared if the initial extracts were first treated with highly purified myrosinase. Peaks A, B, and C contained no significant inducer activity, and cyclocondensation assay of myrosinase hydrolysates showed that only Peaks A and C produced significant quantities of isothiocyanates, accounting for all the inducer activity. See Zhang et al., Anal. Biochem. 205: 100-107 (1992). Peak B was not further characterized. Peaks A and C were eluted from HPLC as TOA salts but required conversion to ammonium salts for successful mass spectroscopy, NMR and bioassay. The pure peak materials were dried in a vacuum centrifuge, redissolved in aqueous 20 mm NH₂Cl, and extracted with chloroform to remove excess TOA bromide. The ammonium salts of glucosinolates remained in the aqueous phase, which was then evaporated.

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Identification of Glucosinolates. The ammonium salts of Peaks A and C were characterized by mass spectrometric (a) negative ion Fast Atom and NMR techniques: Bombardment (FAB) on a thioglyerol matrix; this gave values of 436 (Peak A) and 420 (Peak C) amu for the negative molecular ions, and (b) high resolution NMR, as shown in Figure 2, provided unequivocal identification of Peak A is glucoraphanin structure. methylsulfinylbutyl glucosinolate], and Peak C is the [4-methythiobutyl related glucoerucin glucosinolate]. These identifications and purity are also consistent with the inducer potencies; Peaks A and C, after myrosinase hydrolysis had potencies of 36,100 and 4,360 units/ μ mol, respectively, compared with reported CD values of 0.2 μ M (33,333 units/ μ mol) for sulforaphane and 2.3 μ M (2,900 units/ μ mol) for erucin. CD values are the concentrations of a compound required to double the QR specific activity in Hepa 1c1c7 murine hepatoma cells. Since there are no other glucosinolate peaks, and the inducer activity of peak A and C account for the total inducer activity of the extracts, it is

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therefore likely that in this cultivar of broccoli, there are no significant quantities of other inducers, i.e., no indole or hydroxyalkenyl glucosinolates. Further, the isolated compounds are therefore substantially pure.

Example 8

COMPARISON OF AQUEOUS AND ORGANIC SOLVENT TECHNIQUES FOR EXTRACTION OF INDUCER POTENTIAL

Plants were prepared by first surface sterilizing seeds of Brassica oleracea variety italica (broccoli) cultivar Saga, with 70% ethanol followed by 1.3% sodium hypochlorite and 0.001% alconox. The seeds were grown in sterile plastic containers at a density of approximately 8 seeds/cm2 for 72 hours on a 0.7% agar support that did not contain added nutrients. The environment was carefully controlled with broad spectrum fluorescent lighting, humidity, and temperature control (16 hours light, 25°C/8 hours dark, 20°C).

The plants were rapidly and gently collected from the surface of the agar to minimize glucosinolate hydrolysis by endogenous myrosinase released upon plant wounding. A portion of the plants was homogenized with 10 volumes of the DMF/ACN/DMSO solvent at -50°C, as described in Example 1, which dissolves nearly all the nonlignocellulosic plant material. Alternatively, the bulk of the harvested plants was plunged into 5 volumes of boiling water for 3 min to inactivate endogenous extract glucosinolates myrosinase and to The cooled mixture was homogenized, isothiocyanates. centrifuged, and the supernant fluid was stored at -20°C.

Inducer potential of plant extracts, prepared by the two methods described above, was determined by the microtiter plate bioassay as described above. inducer potentials in an average of 5 preparations were 702,000 (DMF/ACN/DMSO extracts) and 505,000 (aqueous extracts) units/g fresh weight of sprouts.

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Spectrophotometric quantitation οf cyclocondensation product of reaction the of isothiocyanates with 1,2-benzenedithiole was carried out as described in Zhang et al., Anal. Biochem. 205: 100-107 (1992). Glucosinolates were rapidly converted to isothiocyanates after addition of myrosinase. About 6% of the total hot water extractable material [dissolved solids] consisted of glucosinolates. These results demonstrate that (a) isothiocyanate levels in the crude plant extracts are extremely low; (b) myrosinase rapidly converts abundant glucosinolates to isothiocyanates; (c) hot water extraction releases over 70% of the inducer activity extractable with a triple solvent mixture permitting recovery of most of the biological activity in a preparation that is safe for human consumption; and (d) over 95% of the inducing potential in the intact plant is present as glucosinolates and therefore no other inducers are present in biologically significant quantities.

Example 9

DEVELOPMENTAL REGULATION OF GLUCOSINOLATE PRODUCTION

Preliminary experiments in which field grown broccoli (cultivar DeCicco) was harvested at sequential time points from the same field indicated that on a fresh weight basis, inducer potential declined from the early vegetative stage through commercial harvest, but appeared to increase at late harvest (onset of flowering). These data suggested that inducer potential might be highest in seeds. Subsequent studies have shown that when seeds of 8 broccoli cultivars were surface sterilized and grown under gnotobiotic conditions, Phase 2 enzyme-inducer potential was highest in seeds and declined progressively (on a fresh weight basis) over time throughout the first 14 days of seedling growth.

Expressed on a per plant basis, however, activity remained constant over this period, suggesting that at

this early stage of growth there was no net synthesis of glucosinolates. However, when the glucosinolate profiles of market stage broccoli heads and 3 day old sprouts (cultivar Emperor) were compared, there was a profound difference in the apparent glucosinolate compositions of these plants.

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Sprouts were prepared by first surface sterilizing seeds of Brassica oleracea variety italica (broccoli) cultivar Emperor with a 1 minute treatment in 70% ethanol, followed by 15 min in 1.3% sodium hypochlorite with approximately 0.001% Alconox detergent. Seeds were grown in sterile plastic containers at a density of approximately 8 seeds/cm² for 72 hours on a 0.7% agar support that did not contain added nutrients. The environment was carefully controlled; broad spectrum fluorescent lighting, humidity and temperature control (16 hours light, 25°C / 8 hours dark, 20°C).

Plants were rapidly and gently collected from the surface of the agar to minimize glucosinolate hydrolysis by endogenous myrosinase released upon plant wounding. Sprouts [approximately 25 mg fresh wt/sprout], were gently harvested and immediately and rapidly plunged into approximately 3 volumes of boiling water in order to inactivate endogenous myrosinase as well as to extract glucosinolates and isothiocyanates from the plant tissue. Water was returned to a boil and maintained at a rolling boil for 3 min. The sprouts were then strained from the boiled infusion [tea, soup] and the infusion was stored at -20°C until assayed.

Market stage heads were obtained by germinating seeds of the same seedlot in a greenhouse in potting soil, transplanting to an organically managed field in Garrett County, MD and harvested at market stage. Heads were immediately frozen upon harvest, transported to the laboratory on ice and extracts were prepared in an identical fashion to those described above for sprouts

except that approximately 3 gram floret tissue samples were used for extraction.

Inducer potential of plant extracts, prepared as described above, was determined by the microtiter plate bioassay method as described in Example 1. Paired ion chromatography revealed two major peaks, probably glucobrassicin and neo-glucobrassicin, in extracts of market stage heads with similar retention times to glucobrassicin (indole-3-ylmethyl glucosinolate) and neo-(1-methoxyindole-3-ylmethyl glucobrassicin glucosinolate). This observation is consistent with published reports on the glucosinolate composition of paired However, plants. broccoli chromatography under the same conditions of identically prepared extracts of 3-day-old sprouts showed absence of glucobrassicin or neo-glucobrassicin. Additionally, 3day-old sprouts of different broccoli cultivars produce different mixtures of glucosinolates. Accordingly, glucosinolate production is developmentally regulated.

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Example 10

EVALUATION OF ANTICARCINOGENIC ACTIVITIES

OF BROCCOLI SPROUT PREPARATIONS IN THE HUGGINS

DMBA (9,10 DIMETHYL-1,2-BENZANTHRACENE)

MAMMARY TUMOR MODEL

Sprouts were prepared by first surface sterilizing seeds of Brassica oleracea variety italica (broccoli) cultivar Saga with a 1 min treatment in 70% ethanol, followed by 15 min in 1.3% sodium hypochlorite with approximately 0.001% Alconox detergent. Seeds were grown in sterile plastic containers at a density of approximately 8 seeds/cm² for 72 hours on a 0.7% agar support that did not contain added nutrients. The environment was carefully controlled with broad spectrum fluorescent lighting, humidity and temperature control (16 hours light, 25°C / 8 hours dark, 20°C).

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The plants were rapidly and gently collected from the surface of the agar to minimize glucosinolate hydrolysis by endogenous myrosinase released upon plant wounding. A large quantity of sprouts was harvested by immediately and rapidly plunging into approximately 3 volumes of inactivate endogenous boiling water in order to myrosinase, as well as extracting glucosinolates and isothiocyanates from the plant tissue. Water was returned to a boil and maintained at a rolling boil for Sprouts were then strained from the boiled infusion [tea, soup] and the infusion was lyophilized and stored as a dry powder at -20°C [designated Prep A]. Other sprouts, similarly prepared were extracted with boiling water, cooled to 25°C and were amended with a quantity of 7 day old daikon sprouts equivalent to approximately 0.5% of the original fresh weight of broccoli sprouts. This mixture was homogenized using a Brinkman Polytron Homogenizer and incubated at 37°C for 2 hours following which it was filtered through a sintered glass filter, lyophilized as above and stored as a dried powder at -20°C [designated Prep B].

QR inducer activity and inducer potential of plant extracts, prepared as described above, was determined by the microtiter plate bioassay method as described above. The induction of QR activity in preparation A is largely predominantly glucoraphanin, due to glucosinolates; which is the glucosinolate of sulforaphane, but this preparation also contains some glucoerucin, which is the The induction QR sulfide analog of glucoraphanin. activity of preparation B is almost exclusively due to isothiocyanates arising from treatment of glucosinolates with myrosinase.

Female Sprague-Dawley rats received at 35 days of age were randomized; 4 animals per plastic cage. All animals received 10 mg DMBA, by gavage in 1 ml sesame oil, at age 50 days. Sprout preparations (A or B) or vehicle control were given by gavage at 3, 2 & 1 day prior to DMBA, on

the day of DMBA (2 hr prior to the DMBA dose) and on the day following DMBA dosing. The vehicle used was 50% Emulphor 620P / 50% water. Animals were maintained on a semi-purified AIN-76A diet ad libitum from the time of receipt until termination of the experiment (167 days of age).

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ANTICARCINOGENIC ACTIVITIES OF BROCCOLI SPROUT EXTRACTS

IN THE DMBA RAT MAMMARY TUMOR HODEL

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MULTIPLICITY: NUMBER OF TUMORS PER RAT	1.79	1.05	0.55
TOTAL TUBOR NUMBER	34	19	11
NUMBER OF ANTHALS AT TERMINATION	19	18	20
Treathent	DMBA only	324 mg/dose (100 µmol sulforaphane equiv.)	424 mg/dose (100 µmol sulforaphane equiv.)
GROUP	CONTROL	PREPARATION A (Glucosinolate)	PREPARATION B (Isothiocyanate)
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The development of palpable tumors was delayed for as much as 5 weeks by the administration of sprout extracts. Rats treated with either Preparation A or B had significantly fewer tumors than the untreated control, and the multiplicity of tumors (tumors per rat) was significantly lower in the animals receiving Preparations A or B.

Example 11 METABOLISM AND CLEARANCE OF GLUCOSINOLATES IN HUMANS

Two male, non-smoking volunteers ages 35 and 40 years, each in good health, were put on a low vegetable diet in which no green or yellow vegetables, or condiments, mustard, horseradish, tomatoes or papayas were consumed. After 24 hours on such a diet, all urine After 24 hours of was collected in 8 hr aliquots. baseline data, subjects ingested 100 ml of broccoli sprout soup (prepared as below), containing 520 μmol of glucosinolates.

sprouts were prepared by first sterilizing seeds of Brassica oleracea variety italica (broccoli) cultivar Saga with a 1 min treatment in 70% ethanol, followed by 15 min in 1.3% sodium hypochlorite with ca. 0.001% Alconox detergent. Seeds were grown in sterile plastic containers at a density of approximately 8 seeds/cm2 for 72 hours on a 0.7% agar support that did The environment was not contain added nutrients. carefully controlled with broad spectrum fluorescent lighting, humidity and temperature control (16 hours The plants were light, 25°C / 8 hours dark, 20°C). rapidly and gently collected from the surface of the agar to minimize glucosinolate hydrolysis by endogenous myrosinase released upon plant wounding. quantity of sprouts was harvested by immediately and rapidly plunged into approximately 3 volumes of boiling water in order to inactivate endogenous myrosinase as well as to extract glucosinolates and isothiocyanates from the plant tissue. Water was returned to a boil and maintained at a rolling boil for 3 min. Following the boiling step, sprouts were homogenized directly in their infusion water for 1 min using a Brinkman Polytron Homogenizer and the preparations were frozen at -79°C until use.

Inducer potential of plant extracts, prepared as described above, was determined by the microtiter plate bioassay method as described above. Inducer potential is predominantly glucosinolates; due to nearly all glucosinolate the is which glucoraphanin, sulforaphane, but some glucoerucin which is the sulfide analog of glucoraphanin was also present. When converted the addition of purified to isothiocyanates by myrosinase, Phase 2 enzyme-inducing potential was 100,000 units/ml and contained 5.2 μ mol of isothiocyanates per ml, as determined by the cyclocondensation reaction described in Example 7. Thus, the subjects consumed a total of 520 μmol of glucosinolates.

Collection of 8 hour urine samples was continued for an additional 30 hours. Urinary excretion of isothiocyanate conjugates (dithiocarbamates) was monitored using the cyclocondensation reaction as described in Example 7.

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TABLE 5 EXCRETION OF DITHIOCARBAMATES BY TWO SUBJECTS INGESTING 520 MICROMOLES OF GLUCOSINOLATES EXTRACTED FROM SAGA BROCCOLI

5	TIME	CONDITION	SUBJECT 1	SUBJECT 2
TO 620		tion Time ours)	µmol Dithiocarba per 8 hour uring collection	amate
· .	8	baseline	1.4	2.7
	16	baseline	2.1	0.9
10	24	baseline	1.7	5.4
	32	1st 8 hour post-dose	23.2	20.4
5	40	2nd 8 hour post-dose	9.9	36.8
	48	3rd 8 hour post-dose	4.4	14.0
Ω 9 6	56	4th 8 hour post-dose	4.2	4.1
15	Total post- average bas	dose minus eline:	39.8	63.2
		rcent of dose:	6.7%	12.2%

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The two subjects studied metabolically converted a significant fraction of the ingested glucosinolates to the isothiocyanates which were converted to cognate dithiocarbamates and measured in the urine.

Example 12 EFFECTS OF PHYSICAL INTERVENTIONS ON SPROUT GROWTH ON PRODUCTION OF INDUCERS OF QUINONE REDUCTASE

Sprouts were prepared by first surface sterilizing seeds of Raphanus sativum (daikon) by a 1 minute treatment with 70% ethanol, followed by a 15 min treatment with 1.3% sodium hypochlorite with approximately 0.001% Alconox detergent. Seeds were grown

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Treated sprouts were irradiated with germicidal UV light for 0.5 hr on days 5 and 6. Treated sprouts were only half the height of the untreated controls. Plants were harvested on day 7 by rapidly and gently collecting the plants from the surface of the agar to minimize glucosinolate hydrolysis by endogenous myrosinase released upon plant wounding. Sprouts were harvested by immediate and rapid plunging into approximately 10 volumes of DMF/ACN/DMSO (1:1:1) at approximately -50°C in order to inactivate endogenous myrosinase as well as to extract glucosinolates and isothiocyanates. Sprouts were immediately homogenized with a ground glass mortar and pestle and stored at -20°C.

Inducer potential of plant extracts, prepared as described above, was determined by the microtiter plate bioassay method as described above. Inducer potential of the UV-treated sprouts was over three times that of untreated controls. Treatment of sprouts with ultraviolet light therefore increased the Phase 2 enzymeinducer potential of the plant tissue.

Although the foregoing refers to particular preferred embodiments, it will be understood that the present invention is not so limited. It will occur to those of ordinary skill in the art that various modifications may be made to the disclosed embodiments and that such modifications are intended to be within the scope of the present invention, which is defined by the following publications and patent applications All mentioned in this specification are indicative of the

level of skill of those in the art to which the invention pertains.

All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application were specifically and individually indicated incorporated by reference in its entirety.

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- 1, Cruciferous sprouts, with the exception of cabbage, cress, mustard and radish sprouts, harvested prior to the 2-leaf stage.
- The cruciferous sprouts according to claim 1, wherein said sprouts are a Brassica oleracea selected from the group of varieties consisting of acephala, alboglabra, botrytis, costata, gemmifera, gongylodes, medullosa, palmifolia, ramosa, sabauda, italica, sabellica, and selensia.
- The cruciferous sprouts according to claim 2, wherein said sprouts are a Brassica oleracea variety italica.
- The cruciferous sprouts according to claim 1, wherein said sprouts are a Brassica oleracea variety botrytis.
- The cruciferous sprouts adcording to claim 1, wherein said sprouts are a Brassica oleracea variety botrytis subvariety cauliflora.
- The cruciferous sprouts according to claim 1, wherein said sprouts are substantially free of Phase 1 enzyme-inducing potential.
- A non-toxic solvent extract of the cruciferous sprouts according to claim 1.
- The non-toxic solvent extract according to claim 7, wherein said solvent is water.
- The non-toxic solvent extract according to claim 8, further comprising a cruciferous vegetable comprising an active myrosinase enzyme.

- 10. The non-toxic solvent extract according to claim 9, wherein said cruciferous vegetable is of the genus Raphanus.
- 11. A method of increasing the chemoprotective amount of Phase 2\enzymes in a mammal, comprising the step of administering an effective quantity of the cruciferous sprouts according to claim 1.
- 12. Cruciferous sprouts harvested prior to the 2leaf stage, wherein said sprouts have at least 200,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3-days of growth from seeds that produce said sprouts and non-toxic levels of indole their breakdown products glucosinolates and goitrogenic hydroxybutenyl glucosinolates.
- 13. The cruciferous sprouts according to claim 12, wherein said sprouts are a Brassica oleracea selected from the group of varieties consisting of acephala, alboglabra, botrytis, costata, gemmifera, gongylodes, medullosa, palmifolia, ramosa, sabauda, italica, sabellica, and selensia.
- 14. The cruciferous sprouts according to claim 13, wherein said sprouts are \ a Brassica oleracea variety italica.
- 15. The cruciferous sprouts according to claim 13, wherein said sprouts are a Brassica oleracea variety botrytis.
- 16. The cruciferous sprouts according to claim 15, wherein said sprouts are a Brassica oleracea variety botrytis subvariety cauliflora
- 17. A non-toxic solvent extract of the cruciferous sprouts according to claim 12.

- The non-toxic solvent extract according to claim 17, wherein said solvent is water.
- The non-toxic solvent extract according to claim 18, further comprising a cruciferous vegetable comprising an active myrosinase enzyme
- The non-toxic solvent extract according to claim 19, wherein said cruciferous vegetable is of the genus Raphanus.
- 21. A method of preparing a food product rich in glucosinolates, comprising germinating cruciferous seeds, with the exception of cabbage, dress, mustard and radish seeds, and harvesting sprouts prior to the 2-leaf stage, to form a food product comprising a plurality of sprouts.
- 22. The method according to claim 21, wherein said sprouts contain non-toxic levels of indole glucosinolates goitrogenic and breakdown products their and hydroxybutenyl glucosinolates.
- 23. The method according to claim 21, wherein said seeds are a Brassica oleracea selected from the group of varieties consisting of acephala, alboglabra, botrytis, costata, gemnifera, gongylodes, italica, medullosa, palmifolia, ramosa, sabauda, sabellica, and selensia.
- 24. The method according to claim 23, wherein said seeds are Brassica oleracea variety italica.
- 25. The method according to claim 23, wherein said seeds are Brassica oleracea variety botrytis.
- 26. The method according to claim 25, wherein said seeds are Brassica oleracea variety botrytis subvariety cauliflora.

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- 27. A food product rich in glucosinolates made by the process according to claim 21.
- 28 A method of preparing a food product, comprising extracting glucosinolates and isothiocyanates from cruciferous sprouts according to claim 1 with a non-toxic solvent, removing the extracted sprouts from said solvent, and recovering the extracted glucosinolates and isothiocyanates.
- 29. A method of preparing a food product according to claim 28, wherein active myrosinase enzyme is mixed with said cruciferous \sprouts, or said extracted glucosinolates and isothiocyanates, or both said cruciferous sprouts or said extract.
- A method of preparing a food product rich in glucosinolates, comprising derminating cruciferous seeds that produce sprouts having at least 200,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3-days of growth and which contain non-toxic levels of indole glucosinolates and their goitrogenic hydroxybutenyl breakdown products and glucosinolates, and harvesting sprouts prior to the 2-leaf stage to form a food product comprising a plurality of sprouts.
- 31. The method according to claim 30, wherein said seeds are a Brassica oleracea selected from the group of varieties consisting of acephala, alboglabra, botrytis, costata, gemmifera, gongylodes) italica, medullosa, palmifolia, ramosa, sabauda, sabellica, and selensia.
- The method according to claim 31, wherein said seeds are Brassica oleracea variety italica.
- 33. The method according to claim 31, wherein said seeds are Brassica oleracea variety botrytis.

- 34. The method according to claim 33, wherein said seeds are Brassica oleracea variety botrytis subvariety cauliflora.
- 35. A food product kich in glucosinolates, made by the process according to dlaim 30.

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3 A method of preparing a food product, comprising introducing cruciferous seeds, wherein said seeds produce sprouts having at least 200,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3-days of growth and non-toxic levels of indole and their breakdown products glucosinolates goitrogenic hydroxybutenyl glucosinolates, into another edible ingredient

A method of preparing a food product, comprising extracting glucosinolates and isothiocyanates with a nontoxic solvent and isothiocyanantes from cruciferous seeds, sprouts, plants or plant parts wherein seeds that produce said sprouts, plant, or plant parts, have at least 200,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3-days of growth and wherein said seeds, sprouts, plants or plant parts have non-toxic levels of indole glucosinolates and their breakdown products and goitrogenic hydroxybutenyl recovering the extracted glucosimolates, and glucosinolates and isothiocyanates.

- 38. A method of preparing a food product according to claim 37, wherein adtive myrosinase enzyme is mixed with said cruciferous seeds, sprouts or plants; or said extracted glucosinolates and isothiocyanates; or both said cruciferous seeds, sprouts or plants and said extract.
- A method of reducing the level of carcinogens in a mammal, comprising administering to a mammal an

effective amount of cruciferous sprouts, with the exception of cabbage,\cress, mustard and radish sprouts.

- 40. A method of reducing the level of carcinogens in a mammal, comprising administering to a mammal an effective amount of cruciferous sprouts having at least 200,000 units per gram fresh weight of Phase 2 enzymeinducing potential when measured after 3-days of growth from seeds that produce said sprouts and non-toxic levels of indole glucosinolates and their breakdown products and goitrogenic hydroxybutenyl glucosinolates.
- 41./ A method of extracting glucosinolates and isothiccyanates from plant tissue comprising the steps of homogenizing said plant tissue in an excess of a mixture of dimethyl sulfoxide, acetonitrile and dimethylformamide at a temperature sufficient to inactivate myrosinase enzyme activity.
- 42. A food product comprising cruciferous sprouts, with the exception of cabbage, cress, mustard and radish sprouts, harvested prior to the 2-leaf stage, cruciferous seeds; extracts of said sprouts or seeds; or any combination of said sprouts, seeds or extracts.
- 43/ A method of increasing the chemoprotective amount of Phase 2 enzymes in a mammal, comprising the step of administering an effective quantity of the food product according to claim 42.
- A food product comprising cruciferous sprouts harvested prior to the 2-leaf stage, wherein said sprouts have at least 200,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3days of growth from seeds that produce said sprouts and non-toxic levels of indole glucosinolate and goitrogenic glucosinolates; cruciferous seeds; hydroxybutenyl extracts of said sprouts or seeds; or any combination of said sprouts, seeds or extracts.

- 45. A method of \increasing the chemoprotective amount of Phase 2 enzymes in a mammal, comprising the step of administering an effective quantity of the food product according to claim 44.
- 46. Cruciferous sprouts harvested prior to the 2leaf stage, wherein the ratio of monofunctional to bifunctional inducers is at least 20 to 1.
- 47/ A food product supplemented with a purified or partially purified glucosinolate.

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Docket No. 46528/102/JOHO

DECLAR. ION AND POWER OF A'L ORNEY

As a below named inventor, I hereby declare that:

My residence, post office address, and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

CANCER CHEMOPROTECTIVE FOOD PRODUCTS

the specification	of	which	(check	one)
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网	is attached hereto

and was amended on (if applicable). was filed on as Application Serial No.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is known by me to be material to patentability as defined in Title 37, Code of Federal Regulations § 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, § 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN APPLICATION(S)

NUMBER	COUNTRY	DAY/MONTH/YEAR FILED	PRIORITY CLAIMED
<u> </u>			

Figure by claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below and insofar as the subject traiter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose information which is known by me to be material to parentability as defined in Title 37, Code of Federal Regulations § 1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

	APPLICATION SERIAL NO.	FILING DATE	STATUS: PATENTED, PENDING, ABANDONED
E		·	
F			
Ī			

Thereby appoint as my attorneys, with full powers of substitution and revocation, to prosecute this application and transact all business in the Relent and Trademark Office connected therewith: Stephen A. Bent, Reg. No. 29,768; David A. Blumenthal, Reg. No. 26,257; John J. Feldhaus, Reg. No. 23,822; Donald D. Jeffery, Reg. No. 19,980; Eugene M. Lee, Reg. No. 32,039; Peter G. Mack, Reg. No. 26,001; Brian J. MelNamara, Reg. No. 32,789; Sybil Meloy, Reg. No. 22,749; George E. Quillin, Reg. No. 32,792; Colin G. Sandercock, Reg. No. 31,298; Bernhard D. Saxe, Reg. No. 28,665; Richard L. Schwaab, Reg. No. 25,479; Arthur Schwartz, Reg. No. 22,115; Harold C. Wegner, Reg. No. 25,479; Arthur Schwartz, Reg. No. 22,115; Harold C. Wegner, Reg.

Send all correspondence to FOLEY & LARDNER, 3000 K Street, N.W., Suite 500, Washington, DC 20007-5109. Address telephone communications to <u>Bernhard D. Saxe</u> at (202) 672-5300.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of First or Sole Inventor Jed W. FAHEY	Signature of First or Sole Inventor		P/3/95
Residence Address 6704 RIDGE RD., ELDERSHURG, MD	21784	Country of Citizenshi United States	,
Post Office Address 6704 RIDGE PD., ECURIDARE, MD	21784		

Signatures should conform to names as typewritten.

Additional inventors on attached Page 2.

PAGE 2

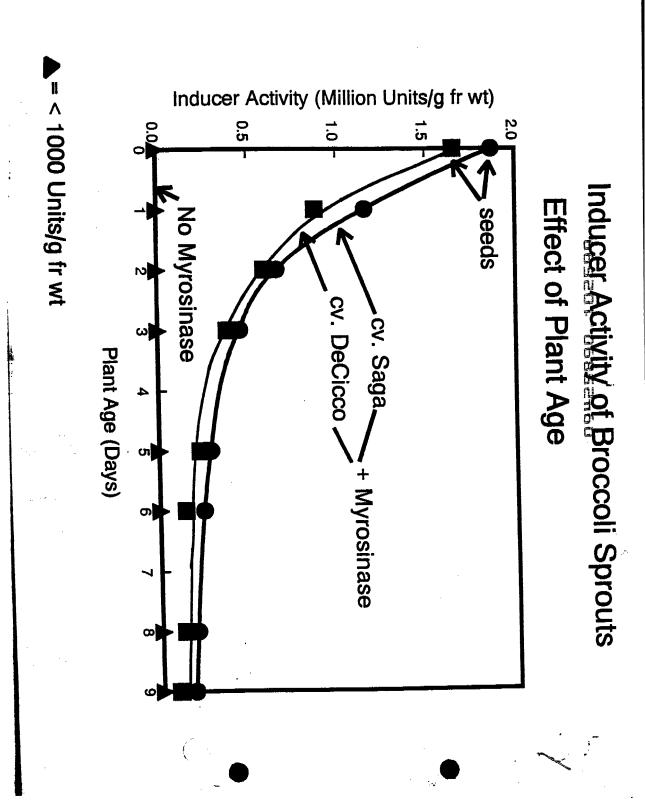
Docket No. 46528/102/JOHO

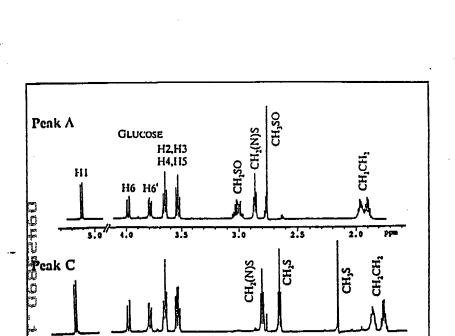
Full Name of Second Inventor Paul TALALAY		of Second Intor	Date 9/13/95
Residence Address 5512 BOXHILL LA	LNE BALTIMORE M	Country of Citizens United States	ship
Post Office Address 5512 BOXHILL LA	NE BALTIMORE	MD 21210	e of est

DOWNERS TAREOR

		Patentee:		A three Toles N	T- 46528/1	02/JOHO	
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				to act on behalf of the		nization identified be	elow:
	NAME	OF ORGANIZA	TION: Johns	Hopkins School of Me 24 E. Monument Stree	dicine	Paltimore MD 2120	75
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Ø		the specification	filed herewith				
C		application seria	i no	, filed		·	
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High Resolution NMR (600 MHz) in D₂O. Note: chirality of SO in Peak A induces multiplet for CH₂SO (Peak A), not observed for CH₂S (Peak C).

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Figure 2

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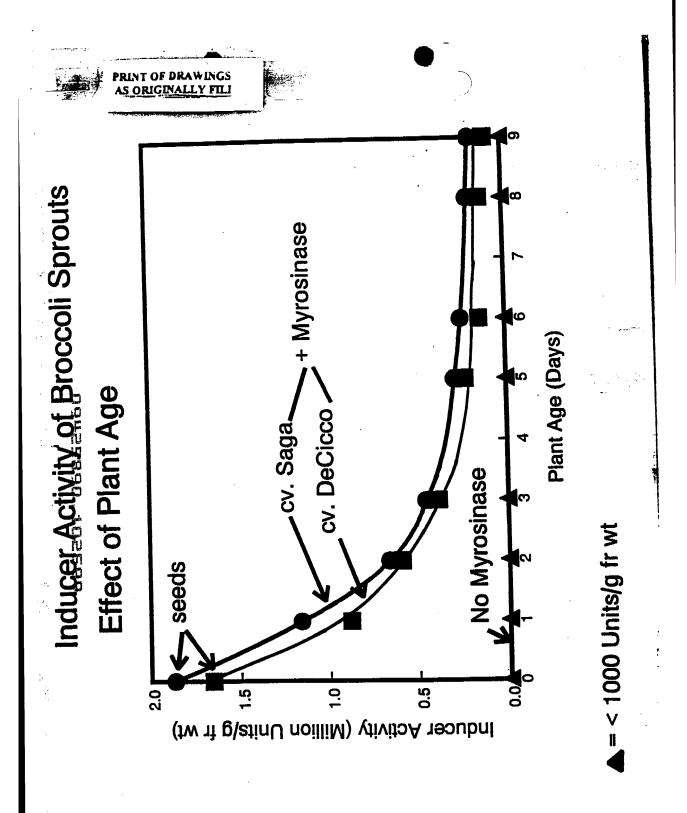
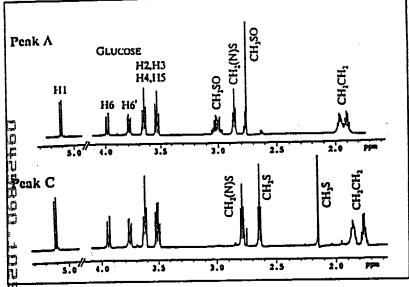


Figure (





Filed 05/09/2008



High Resolution NMR (600 MHz) in D₂O. Note: chirality of SO in Peak A induces multiplet for CH₂SO (Peak A), not observed for CH₂S (Peak C).

Figure 2

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MARCH 14, 1996

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B SAXE FOLEY & LARDNER P.O. BOX 25696 3000 K STREET, N.W., SUITE 500 WASHINGTON, D.C. 20007-5109

UNITED STATES DEPARTMENT OF COMMERCE Patent and Trademark Office
ASSISTANT SECRETARY AND COMMISSIONER OF PATENTS AND TRADEMARKS Washington, D.C. 20231



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PLEASE REVIEW ALL INFORMATION CONTAINED ON THIS NOTICE. THE INFORMATION CONTAINED ON THIS RECORDATION NOTICE REFLECTS THE DATA PRESENT IN THE PATENT AND TRADEMARK ASSIGNMENT SYSTEM. IF YOU SHOULD FIND ANY ERRORS OR HAVE QUESTIONS CONCERNING THIS NOTICE, YOU MAY CONTACT THE EMPLOYER WHOSE NAME APPEARS ON THIS NOTICE AT 703-308-9723. PLEASE SEND REQUEST FOR CORRECTION TO: U.S. PATENT AND TRADEMARK OFFICE, ASSIGNMENT DIVISION, BOX ASSIGNMENTS, NORTH TOWER BUILDING, SUITE 10C35, WASHINGTON, D.C. 20231.

RECORDATION DATE: 09/15/1995

REEL/FRAME: 7694/0746 NUMBER OF PAGES: 2

BRIEF: ASSIGNMENT OF ASSIGNOR'S INTEREST (SEE DOCUMENT FOR DETAILS).

ASSIGNOR:

FAHEY, JED W.

DOC DATE: 09/13/1995

ASSIGNOR:

TALALAY, PAUL

DOC DATE: 09/13/1995

ASSIGNEE:

JOHNS HOPKINS SCHOOL OF MEDICINE 2024 E. MONUMENT STREET, SUITE 2-100 BALTIMORE, MARYLAND 21205

SERIAL NUMBER: 08528858 PATENT NUMBER:

FILING DATE: 09/15/1995

ISSUE DATE:

SEDLEY PYNE, EXAMINER ASSIGNMENT DIVISION OFFICE OF PUBLIC RECORDS

Sembard D. Sens Name of Person Signing

4,97 15:13 No.006 JHU-TECHNOLOGY LICENSING TEL: 410-955-1245 11-21-1995 CMB No. 0651-0011 (exp. 4/9 untimus uniginal documents or copy To the Honorable Commissio 2. Name and address of receiving party(les): 1. Name of conveying partylisal Name: Johns Hooldes School of Medicine Jed W. FAHEY, Parl TALALAY imternal Address: Additional name(s) of conveying party(los) attached? No Street Address: 2024 E. Monument Street, Suits 2-100 3. Nature of conveyance: City: Beltimore, State: MD ZIP: 21205 Merger XX Assignment Change of Name Security Agreement Other Additional name(s) & address(es) attached? No Execution Date: 09-13-95 4. Application number(s) or patent number(s): If this document is being filed together with a new application, the execution date of the application is: 98-15-95 B. Patent No.(s) A. Patent Application No.(s) Additional numbers attached? No 6. Total number of applications and patents 5. Name and address of party to whom correspondence involved: 1 concern vg document should be mailed: 7. Total fee (37 C.F.R. § 3.41). \$40.00 Name: | OLEY & LARDNER - Attn: B. Saxe Internal Address: P.O. Box 25696 XX Enclosed Authorized to be charged to deposit account Street Address: 3000 K Street, N.W., Suite 500 City: Washington, D.C. ZIP: 20007-5109 8. Deposit account number: (Attach duplicate copy of this page if paying by deposit account) DO NOT USE THIS SPACE 110 HG 10/11/95 08528858 40.00 68 Statement and signature. and any ettached copy is a true copy of the original To the best of my knowledge and belief, the foregoing information is true and comdocument. September 15, 1996

> Meil documents to be recorded with required cover sheet information to: Commissioner of Patents & Trademarks, Box Assignments Washington, D.C. 20231

Total number of pages including cover sheet, attachments, and document: 2

INMENT - WORLDY I

For good and valuable consideration, the receipt and sufficiency of which are hereby acknowledged, each undersigned inventor has sold and assigned, and by these presents hereby sells and assigns, unto

JOHNS HOPKINS SCHOOL OF MEDICINE

its successors and assigns, the entire right, title and interest, so far as concerns the United States and the Territories and Possessions thereof and all foreign countries in and to the invention in

CANCER CHEMOPROTECTIVE FOOD PRODUCTS

XX	executed con-	currently herewith	
_	executed on _		
	Serial No.	filed _	

as set forth in his United States Patent Application

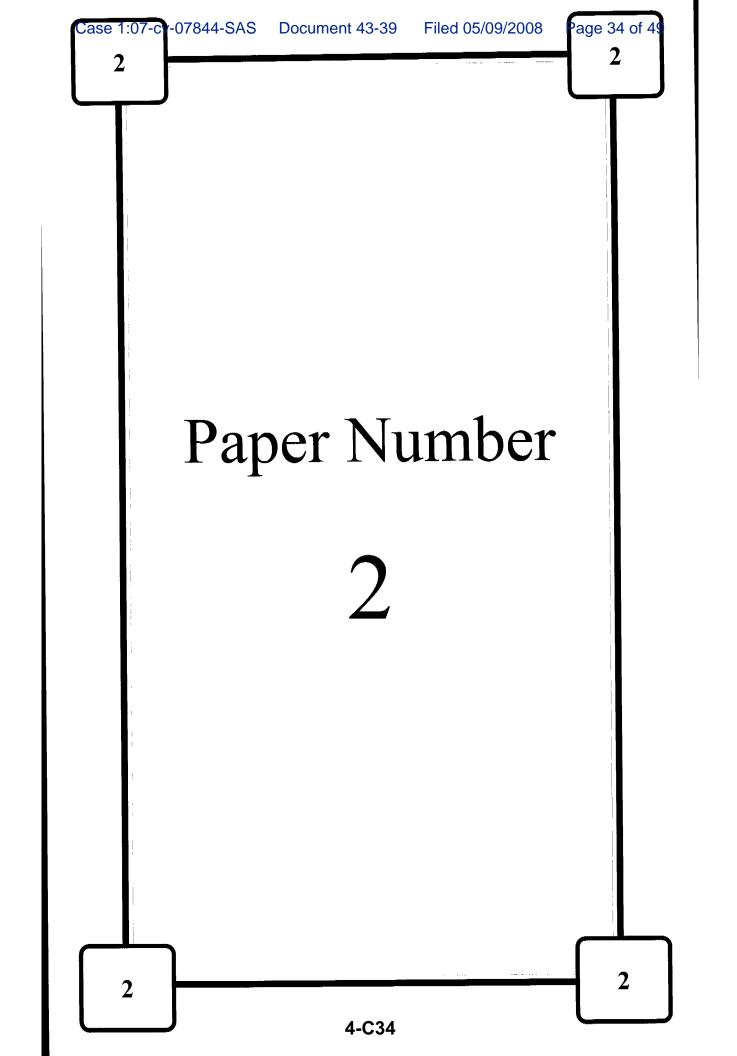
said application for United States Letters Patent, including all divisional, renewal, substitute, continuation and Convention applications based in whole or in part upon said inventions or upon said applications, and any and all Letters Patent and reissues and extensions of Letters Patent granted for said inventions or upon said applications and every priority right that is or may be predicated upon or arise from said inventions, said applications, and said Letters Patent; said Assignee being hereby authorized to file patent applications in any or all countries on any or all said inventions in the name of the undersigned or in the name of said Assignee or otherwise as said Assignee may deem advisable, under the International Convention or otherwise; the Commissioner of Patents and Trademarks of the United States of America being hereby authorized to issue or transfer all said Letters Patent to said Assignee in accordance herewith; this assignment being under covenant, not only that full power to make the same is had by the undersigned, but also that such assigned right is not encumbered by any grant, license, or other right theretofore given, and that the undersigned will do all acts reasonably serving to ensure that the said inventions, patent applications and Letters Patent shall be held and enjoyed by said Assignee as fully and entirely as the same could have been held and enjoyed by the undersigned if this assignment had not been made, and particularly to execute and deliver to said Assignee all lawful documents including petitions, specifications, oaths, assignments, invention disclaimers, and lawful affidavits in form and substance which may be requested by said Assignee, to furnish said Assignee with all facts relating to said inventions or the history thereof and any and all documents, photographs, models, samples or other physical exhibits which may be of said inventions, and to testify in any proceedings relating to said inventions, patent applications and Letters Patent.

The undersigned hereby grant the firm of FOLEY & LARDNER the power to insert in this Assignment any further identification which may be necessary or desirable to comply with the rules of the U.S. Patent and Trademark Office for recordation of this Assignment.

NAMES AND SIGNATURES OF INVENTORS							
Name: Jed W. FAHEY Signature: Sed (i) Select 1/13/95							
Name:Paul TALALAY	Signature: Faul Tala Cay	Date: 9//3/95					
Name:	Signature: /	Date:					
NAME	S AND SIGNATURES OF WITNESSES						
Name: RUTH DILLINGER	Signature: Becch Williams	Date: 9//3/95					
Name: SHARON KERRY	Signature: X - Kum	Date: 9-13-95					

Note: Prima facie evidence of execution may optionally be obtained by execution of this document before a U.S. Consul or before a local officer authorized to administer oaths whose authority is proved by a certificate from a U.S. Consul.

(PT-407:4/90(1)(modified)



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No. 046528/0121

In re patent application of

Jed FAHEY et al.

Serial No. Unassigned

Filed: October 25, 1999

For:

CANCER CHEMOPROTECTIVE FOOD PRODUCTS

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents Washington, D.C. 20231

Sir:

Prior to examination of the above-identified application, Applicants respectfully request that the following amendments be entered into the application:

IN THE CLAIMS:

Kindly cancel claims 1-35 and 39-47 without prejudice or disclaimer.

REMARKS

Claims 36-38 are now pending. Claims 1-35 and 39-47 have been canceled. Entry of the foregoing amendment prior to examination is respectfully requested.

Respectfully submitted,

October 25, 1999

Registration No. 35,792

FOLEY & LARDNER 3000 K Street, N.W. Suite 500 Washington, D.C. 20007-5109

Tel: (202) 672-5300



Law Offices
FOLEY & LARDNER
Suite 500
3000 K Street, N.W.
Washington, DC 20007-5109
(202) 672-5300



TO: Assistant Commissioner for Patents Box Patent Applications Washington D.C. 20231

Attorney Docket No.046585/0121

Copies of IDS Citations

UTILITY PATENT APPLICATION TRANSMITTAL (new nonprovisional applications under 37 CFR 1.53(b))

Transmitted herewith for filing is the patent application of:

INVENTOR(S): Jed W. FAHEY and Paul TALALAY

TITLE: CANCER CHEMOPROTECTIVE FOOD PRODUCTS

In connection with this application, the following are enclosed:

APPLICATION ELEMENTS: Specification - 51 TOTAL PAGES T XX FN (preferred arrangement:) -Descriptive Title of the Invention V. -Cross Reference to Related Applications Ţ -Statement Regard Fed sponsored R&D -Reference to Microfiche Appendix đ -Background of the Invention -Brief Summary of the Invention -Brief Description of the Drawings (if filed) ÷ -Detailed Description -Claim(s) -Abstract of the Disclosure XX Drawings - Total Sheets 2 C Declaration and Power of Attorney - Total Sheets 2 XX __ Newly executed (original or copy) XX Copy from a prior application (37 CFR 1.63(d)) (relates to continuation/divisional boxes completed) - NOTE: Box below $\underline{\texttt{DELETION}} \ \ \textbf{OF} \ \ \underline{\texttt{INVENTOR}(S)} \ \ \textbf{-} \ \ \\ \textbf{Signed} \ \ \textbf{statement} \ \ \textbf{attached} \ \ \textbf{deleting} \ \ \textbf{inventor}(s)$ named in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b). Incorporation By Reference (useable if copy of prior application XX Declaration being submitted) The entire disclosure of the prior application, from which a COPY of the oath or declaration is supplied as noted above, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein. Microfiche Computer Program (Appendix) Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary) Computer Readable Copy Paper Copy (identical to computer copy) Statement verifying identify of above copies ACCOMPANYING APPLICATION PARTS Assignment Papers (cover sheet & document(s))

37 CFR 3.73(b) Statement (when there is an assignee)

Information Disclosure Statement (IDS) with PTO-1449.

English Translation Document (if applicable)

XX Preliminary Amendment

XX Return Receipt Postcard (MPEP 503)

XX Amend the specification by inserting before the first line the following sentence: /-This application is a divisional of prior of application Serial No. 09/118,867, filed July 20, 1998, now pending; which is a divisional of 08/840,234, filed April 11, 1997, now U.S. Patent 5,968,567, issued 10-19-99.--

CORRESPONDENCE ADDRESS:

Foley & Lardner Address noted above.

U.S. Patent 5,968,567, issued 10-19-99.

Telephone: 202-672-5300 Fax Number: 202-672-5399

100	PER CALCIDATIO	NS: (Small entit	y fees indicate	d in parenth	eses.)
		(2) Number Filed	(3) Number Extra	(4) Rate	(5) Basic Fee \$760 (\$380)
	Total Claims	3 - 20 =	0	x \$18 (x \$9)	0
1000	Independent	2 - 3 =	0	x \$78 (x \$39)	0
	Multiple Dependent Claims			\$260 (\$130)	0
	Assignment Re	ecording Fee per	property	\$40	0
		der 37 C.F.R. 1.1		\$130 (\$65)	0
				TOTAL FEE:	\$380.00

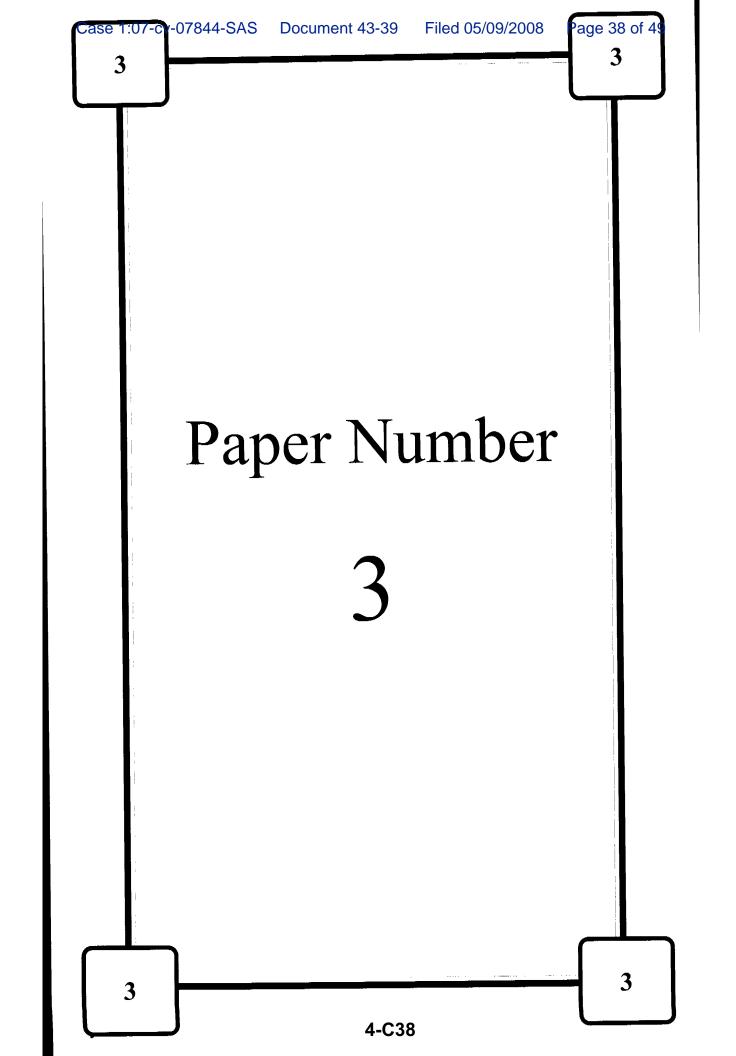
METHOD OF PAYMENT:

A check in the amount of the above TOTAL FEE is attached. If payment by check is NOT enclosed, it is requested that the Patent and Trademark Office advise the undersigned of the period of time within which to file the TOTAL FEE. If payment enclosed, this amount is believed to be correct; however, the Commissioner is hereby authorized to charge any deficiency or credit any overpayment to Deposit Account No. 19-0741.

Respectfully submitted,

Date: October 25, 1999 Docket No.: 046585/0121

Richard C. Peet Reg. No. 35,792



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No. 046585/0121

Page 39 of 49

In re patent application of

Jed FAHEY et al.

Serial No. 09/425,890

Filed: October 25, 1999

For: CANCER CHEMOPROTECTIVE FOOD PRODUCTS

Group Art Unit: 1761 Examiner: Unknown

INFORMATION DISCLOSURE STATEMENT UNDER 37 C.F.R. §1.56

Assistant Commissioner for Patents Washington, D.C. 20231

Sir:

Submitted herewith on a modified Form PTO-1449 is a listing of documents known to applicants in order to comply with applicant's duty of disclosure pursuant to 37 C.F.R. §1.56.

The documents listed are cited in the parent application. The listed documents include documents that became known to applicant incident to a suit for infringement of U.S. Patent No. 5,725,895 filed in the District Court of Delaware. U.S. Patent No 5,725,895 is not related to the present application. However, applicants note that the present application and U.S. Patent No. 5,725,895 have an inventor in common and relate, generally, to the same technical field. Accordingly, out of an abundance of caution, and in compliance with the duty of disclosure, applicant hereby brings these documents to the attention of the Patent Office

The accompanying Form PTO-1449 lists several papers and publications that were provided during the course of discovery in the infringement suit. In addition, the defendants have recently filed a request for reexamination of U.S. Patent No. 5,725,895 citing several of the listed papers and publications.

Applicant believes that the documents listed in the accompanying Form PTO-1449 do not adversely impact the patentiality of the claims of the above-captioned application. However, out of an abundance of caution, and in compliance with the duty of disclosure, applicant hereby brings these documents to the attention of the Patent Office.

In the course of the infringement suit related to U.S. Patent No. 5,725,895, the defendants also have lodged several affirmative defenses and counterclaims, including (1) invalidity and unenforceability for failure to comply with the provisions of 35 U.S.C. §§ 101, 102, 103, and 112, (2) breach of the duty to disclose material information, and (3) patent misuse. The defendants' "Answer, Affirmative Defenses and Counterclaim," which contains these allegations, also is listed on the accompanying Form 1449.

Applicant believes that the foregoing affirmative defenses and counterclaims are without merit. However, out of an abundance of caution, and in compliance with the duty of disclosure, applicant hereby brings these documents to the attention of the Patent Office.

The submission of any document herewith, which is not a statutory bar, is not intended as an admission that such document constitutes prior art against the claims of the present application or that such document is considered material to patentability as defined in 37 C.F.R. §1.56(b). Applicant does not waive any rights to take any action which would be appropriate to antedate or otherwise remove as a competent reference any document which is determined to be a prima facie prior art reference against the claims of the present application.

CONCISE EXPLANATION OF RELEVANCE OF EACH DOCUMENT

Applicants are submitting herewith on Form PTO-1449, a listing of the documents cited by or submitted to the Patent Office in parent application Serial No. 09/118,867, filed July 20, 1998, which is a divisional application of Serial No. 08/840,234, filed April 11, 1997, now U.S. Patent No. 5,968,567. The relevance of these prior art documents is explained in the parent application.

As provided in 37 C.F.R. §1.98(d), copies of the documents are not being provided since they were previously cited by or submitted to the Patent Office in parent application Serial No. 08/840,234, filed April 11, 1997, which is a divisional application of Serial No. 08/528,858, filed September 15, 1995, now U.S. Patent No. 5,725,895.

Since this Information Disclosure Statement is being filed in compliance with 37 C.F.R. §1.97(b) before mailing of a first Office Action on the merits, no fee is required in connection with its filing.

Applicant respectfully requests that the listed documents be considered by the Examiner and be made of record in the present application and that an initialed copy of Form PTO-1449 be returned in accordance with MPEP §609.

Respectfully submitted,

DANNAY 21 2000

Date

For Richard C. Peet

Registration No. 35,792

FOLEY & LARDNER 3000 K Street, NW, Suite 500 Washington, DC 20007-5109 (202) 672-5300

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		TALEN	T OF COMMERCE	ATTY, DOCKET	-	SERIAL NO.		
Form PTO-144	TRADEMARK OFFICE				0121	09/425,890		
(MODIFIED)		PATENT F.AD TO	(DE(W)	APPLICANT	APPLICANT			
7 INFO	DMATIO	N DISCLOSURE	CITATION		Jed W. FAHI			
	KINALIO	11 0100		FILING DATE		GROUP AR	T UNIT	
	570.	1 20 2000		October 2	5, 1999		1761	
Date Submitte	d to PIU:	January 20, 2000					Sheet 1 of 4	
			U.S. PATI	ENT DOCUMENTS				
EXAMINER INITIAL	REF	DOCUMENT NUMBER	DATE	NAME	CLASS	SUB- CLASS	FILING DATE IF APPROPRIAT	
	A1	5,725,895	3/1998 F	ahey et al.	426	49		
LAW		5,411,986	5/1995 C	Cho et al. PE JOR	514	514.		
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		 		1000	<u> </u>	<u></u>	* :	

FOREIGN PATENT DOCUMENTS

		TO COLUMN TO LIT	01.400	SUB-	TRANSLATION			
	REF	DOCUMENT NUMBER	DATE	COUNTRY	CLASS	CLASS	YES	NO
<u></u>	A8							
	A9							
	A10							
	A11				War at Barran Etc	<u> </u>		

OTHER DOCUMENT(S) (Including Author, Title, Date, Pertinent Pages, Etc.)

	OTH	HER DOCUMENT(S) (Including Author, Title, Date, Pertinent Pages, Etc.)
	A12	The Sproutletter, Number 25, Nov. – Dec. 1984
·AW	A13	"The Sproutletter" May-June 1981, No. 4
	A14	Roy Bruder, Ph.D., Discovering Natural Foods, (including pgs.203-209), Woodbridge Press, 1982.
	A15	Brian R. Clement, Hippocrates Health Program, (including pgs 7-11), Hipprocrates Publications, 1989.
	A16	Jethro Kloss, The Back to Eden Cookbook, pgs. 61-61, Woodbridge Press, 1974.
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	A18	Steve Meyerowitz, Sprout It, One week from Seed to Salad, The Sprouthouse, Inc., (including pgs. 84-85, 120-123), June 1994.
	A19	Steve Meyerowitz, The Complete Guide to Sprouting, Sprouts The Miracle Food, Sproutman Publications, (including pgs. 121-2), May 1998.
	A20	Esther Munroe, Sprouts to Grow and Eat, (including pgs. 2-15), Dec. 1974.
	A21	Jean Hewitt, The New York Times "New Natural Foods Cookbook:, Avon Books, pgs. 200-203, 1982.
	A22	Martha H. Oliver, Add a Few Sprouts To Eat Better for Less Money, Pivot Original Health Books, (including pgs. 52-53, 118-119), 1975.
	A23	James C. Schmidt, Horticulture Facts, "Growing Sprouts Indoors", (Rev. 4/81).
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(MODIFIED)			ADEMARK OFFICE	046585/0121	09/425,890		
Ď			i	APPLICANT			
NFO INFO	RMATIO	N DISCLOSURE	CITATION	Jed W.	FAHEY et al.		
			i	FILING DATE	GROUP ART UNIT		
Date Submitte	d to PTO:	January 20, 2000		October 25, 1999	1761		
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	;		Schocken Books, 19	981.			
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	A32		Ann Wigmore, "The Hippocrates Diet and Health Program", Avery Publications, 1984.				
	A33	OF	Sprouting Publications Oahspe Foundation, Health and Sprouting Supplies, date N.A.				
	A34	1/()	Sproutletter, #41, Su				
	A35	M 2 1 2000	The Sproutletter, Nu.	ımber 27, March-April 1985.			
	A26	1 1/4/	Steve Meverowitz G	Proving Vegetables Indoors" 198	3.		

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-	A37	(A) -(1)	The Sproutletter, Number 24, SeptOct. 1984.		
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	A39		The Sproutletter, Number 28, May-June 1985.		
1	A40		The Sproutletter, Number 26, Jan-Feb 1985.		
	A41		Sprouting Publications, Health and Sprouting Supplies, date NA.		
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	A43		Sproutletter, #40, Spring, 1989.		
	A44		The Sproutletter, Number 32, Summer., dode N.A.		
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	A47		Sproutletter, #39, Fall, 1988.		
	A48		Sproutletter, #43, May/June 1990.		
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VIW	A50		Sprouting Publications Health and Sprouting Supplies. , & & C C . A .		
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ugu	A52		The Sproutletter, A newletter of useful and unusual information on sprouts, raw foods an nutrition, dot w.k.		
1	A53		The Sproutletter, #31, Winter, Lake W.A.		
	A54		Deirdre Purdy, ed., The Summer Kitchen, A Farmers' Market Cookbook, 1981.		
	A55		Viktoras Kulvinskas, M.S. Co-Director Hippocrates Health Institute, "Love Your Body or how to be a live food lover", 1974.		
	A56		The Sprout House Article from Newspaper, date N.A.		
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	A58		Steve Meyerowitz, Indoor Vegetable Kit, The Sprout House, んぱん なん		
VAW.	A59	1	The Sprout House Newsletter, Issue #15, August, 1992.		

orm PTO-144			NT OF COMMERCE ADEMARK OFFICE	046585/0121	SERIAL NO. 09/425,890	
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\ Data Submitte	d to PTO:	January 20, 2000	NW 5 1 SOUR SE	October 25, 1999	1761	
Jale Gubrille	A60		Springer substice	Seeds for Sprouting 100% Organ	Sheet 3 o ically Grown Order Form.	
	A61		-Complaint for Patent	Infringement (Brassica Protection	n Products, LLC v. The Sproutma	
		***	Inc. dated Geptembe	r-20, 1999. r, Affirmative Defenses and Cour	toroloime dated June 28, 1999	
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		pages 20-21, 85		C)		
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		Posner et al., "Design and Synthesis of Bifunctional Isothiocyanate Analogs of Sulforaphane				
	A71	Correlation between structure and potency as Inducers of Anticarcinogenic Detoxication Enzymes",				
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		Prochaska et al.	, "Regulatory Mechani	sms of Monofunctional and Bifun	ctional Anticarcinogenic Enzyme	
	A75			earch Vol. 48, pp. 4776-4782, Se		
					se from Cells Cultured in Microtite	
	A76		·	inogenic Enzyme Inducers", ana		
	7.70	pp. 328-336, 198			,	
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	A77		.) pp. 1166 s-1170s, 19		G. G. GVIGW 17 W. V. V. W.	
		i ivuu oo (suppi	., pp. 1100 8-11/08. T	<i>00</i> →.		
		Dunatar - 4 - 1 "			oxity carcinogens", Proc. Natl. Ad	

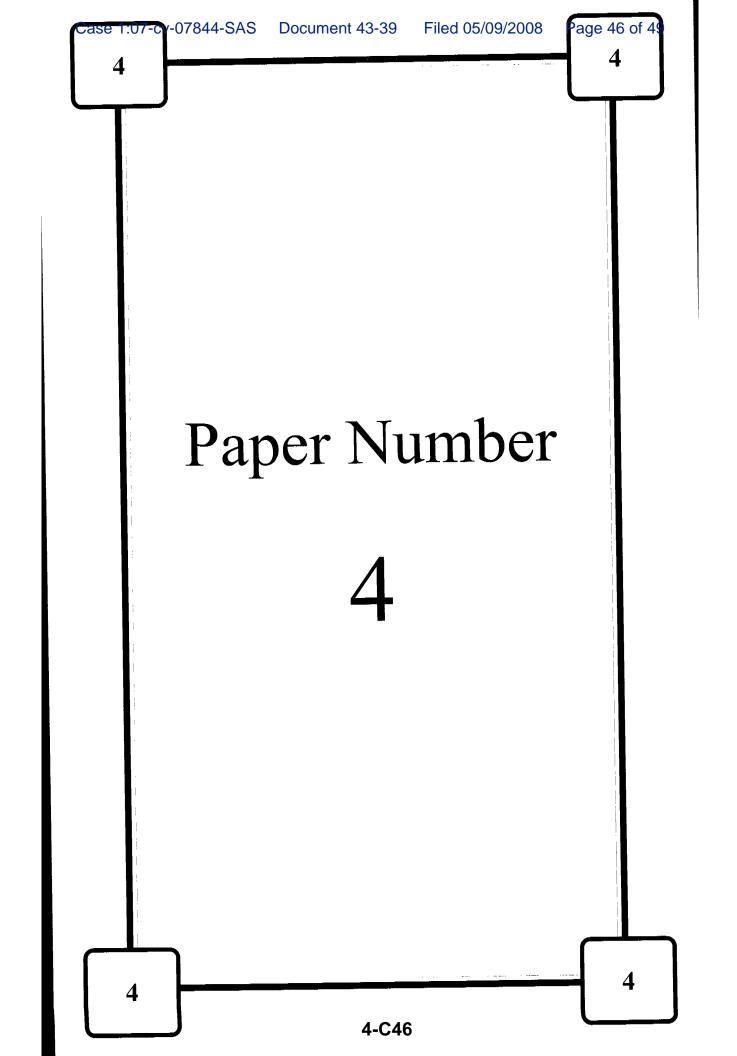
Sheet 4 of 4

Form PTO-1449 U.S. DEF TMENT OF COMMERCE	ATTY, DOCKET	SERIAL NO. 09/425.890	
(MODIFIED) PATENT AND TRADEMARK OFFICE	046585/0121 APPLICANT	09/425,690	
INFORMATION DISCLOSURE CITATION	Jed W. FAHEY et al.		
IN OKUMATION BIOGET	FILING DATE	GROUP ART UNIT	
Date Submitted to PTO: January 20, 2000	October 25, 1999	1761	
Date Continues to the same of		Sheet 4 of	

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		Prestera et al., "The Electrophile Counterattack Response: Protection Against Neoplasia and Toxicity",		
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		"Protocols", Cancer Chemotherapy Reports (Part 2) Vol. 2, No.1, pp. 135-140, April 1971.		
		Polasa et al., "Cancer preventive properties of varieties of Brassica oleracea: A review Source",		
	A83	American Journal of Clinical Nutrition 59 (5 Suppl), 1994.		
	A84	Patent Abstract of Japan Sect. No. 305, Vol. 9, No. 2371, p.2, September 1985.		
		Barrett et al., "Protective Effect of Cruciferous Seed Meals Against Mouse Colon Cancer", Cereal		
JAN	A85	Foods World 613, Vol. 39, No. 8, pp. 613, August 1994.		
XAMINER	<u> </u>	DATE CONSIDERED		

EXAMINER: Initial if citation considered, whether or not citation is in conformance with MPEP 609; Draw line through citation if not in conformance and not considered. Include any copy of this form with next communication to applicant.





IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No. 046585/0121

In re patent application of:

Jed FAHEY et al.

Serial No.: 09/425,890

Filed: October 25, 1999

Group: Unassigned

Examiner: Unassigned

FOR: CANCER CHEMOPROTECTIVE FOOD PRODUCTS

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents Washington, DC 20231

TC 1700 MAIL ROOM

Sir:

Prior to examination of the above-identified application, Applicants respectfully request that the following amendments be entered into the application:

IN THE CLAIMS:

Please add the following new claims:

A method of preparing a human food product comprising cruciferous seeds, Eur cal flour made from the cruciferous seeds, or a combination thereof, wherein the cruciferous seeds or flour contain high Phase 2 enzyme-inducing potential, comprising the steps of:

> selecting eruciferous seeds which produce sprouts that contain at least 200,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3-days of growth, and

preparing a food product from the selected cruciferous seeds. (b)

The method of claim 48, wherein the selected cruciferous seeds produce sprouts that contain at least 300,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3-days of growth.

The method of claim 48, wherein the selected cruciferous seeds produce sprouts that contain at least 400,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3-days of growth.

The method of claim 48, wherein the selected cruciferous seeds produce sprouts that contain at least 500,000 units per gram fresh weight of Phase 2 enzyme-inducing potential when measured after 3-days of growth.--

REMARKS

The foregoing amendments add new claims 48-51 to the present application.

Applicants respectfully request that these amendments be made to the present application prior to examination.

Sérial No. Unassigned

Attorney Docket No.: 046585/121

Upon entry of the foregoing amendments, claims 36-38 and 48-51 are presented for examination. Claims 1-35 and 39-47 are canceled without prejudice or disclaimer.

Respectfully submitted,

November 30, 1999

Date

Richard C. Peet Reg. No. 35,792

Foley & Lardner 3000 K Street, NW Washington Harbour, Suite 500 Washington, DC 20007

Tel: (202) 672-5300 Fax: (202) 672-5399

Should additional fees be necessary in connection with the filing of this paper, or if a petition for extension of time is required for timely acceptance of same, the Commissioner is hereby authorized to charge Deposit Account No. 19-0741 for any such fees.